

Immune Cell Function Assay

(20456)

Medical Benefit		Effective Date: 04/01/12	Next Review Date: 01/15
Preauthorization	No	Review Dates : 09/10, 09/11, 01/12, 01/13, 01/14	

The following Protocol contains medical necessity criteria that apply for this service. It is applicable to Medicare Advantage products unless separate Medicare Advantage criteria are indicated. If the criteria are not met, reimbursement will be denied and the patient cannot be billed. Preauthorization is not required but is recommended if, despite this Protocol position, you feel this service is medically necessary; supporting documentation must be submitted to Utilization Management. Please note that payment for covered services is subject to eligibility and the limitations noted in the patient's contract at the time the services are rendered.

Description

Careful monitoring of lifelong immunosuppression is required to ensure long-term viability of solid organ allografts without incurring an increased risk of infection. The monitoring of immunosuppression parameters attempts to balance the dual risks of rejection and infection. It is proposed that individual immune profiles, such as an immune cell function assay, will help assess the immune function of the transplant recipient and individualize the immunosuppressive therapy.

Background

Currently, immunosuppression is determined by testing for clinical toxicity (e.g., leukopenia, renal failure) and by therapeutic drug monitoring (TDM) when available. However, drug levels are not a surrogate for overall drug distribution or efficacy because pharmacokinetics often differ among individuals due to clinical factors such as underlying diagnosis, age, gender, and race; circulating drug levels may not reflect the drug concentration in relevant tissues; and levels of an individual immunosuppressant drug may not reflect the cumulative effect of other concomitant immunosuppressants. The main value of TDM is the avoidance of toxic levels and monitoring patient compliance. Further, the appropriate level of immunosuppression may vary from person to person. Individual immune profiles, such as an immune cell function assay, could support clinical decision making and help to manage the risk of infection from excess immunosuppression and the risk of rejection from inadequate immunosuppression in immunosuppressed patients.

Regulatory Status

ImmuKnow® (Cylex, Columbia, MD) is an immune cell function assay cleared for marketing by the U.S. Food and Drug Administration (FDA) in April 2002 to detect cell-mediated immunity (CMI) in an immunosuppressed patient population. The assay measures the concentration of adenosine triphosphate (ATP) in whole blood following a 15- to 18-hour incubation with the mitogenic stimulant phytohemagglutinin (PHA). In cells that respond to stimulation, increased ATP synthesis occurs during incubation. Concurrently, whole blood is incubated in the absence of stimulant for the purpose of assessing basal ATP activity. CD4+ T lymphocytes are immunoselected from both samples using anti-CD4 monoclonal antibody-coated magnetic particles. After washing the selected CD4+ cells on a magnet tray, a lysis reagent is added to release intracellular ATP. A luminescence reagent added to the released ATP produces light measured by a luminometer, which is proportional to the concentration of ATP. The characterization of the cellular immune response of a specimen is made by comparing the ATP concentration for that specimen to fixed ATP level ranges.

Last Review Date: 01/14

In April 2002, Cylex obtained 510(k) clearance from the FDA to market the Immune Cell Function Assay based on substantial equivalence to two flow cytometry reagents ("predicate devices") manufactured by Becton Dickinson, the TriTest™ CD4 FITC/CD8 PE/CD3 PerCP Reagent and the MultiTest™ CD3 FITC/CD8 PE/CD45 PerCP/CD4 APC Reagent. These reagents are used to determine CD4+ T-lymphocyte counts in immunocompromised patients. The FDA-indicated use of the Cylex Immune Cell Function Assay is for the detection of cell-mediated immunity in an immunosuppressed population. A subsequent 510(k) marketing clearance for a device modification was issued by the FDA for this assay in 2010. There were no changes to the indications or intended use. (1)

Policy (Formerly Corporate Medical Guideline)

Use of the immune cell function assay to monitor and predict immune function after solid organ transplantation is considered **investigational**.

Use of the immune cell function assay to monitor and predict immune function after hematopoietic stem cell transplantation is considered **investigational**.

Use of the immune cell function assay for all other indications is considered investigational.

Services that are the subject of a clinical trial do not meet our Technology Assessment Protocol criteria and are considered investigational. For explanation of experimental and investigational, please refer to the Technology Assessment Protocol.

It is expected that only appropriate and medically necessary services will be rendered. We reserve the right to conduct prepayment and postpayment reviews to assess the medical appropriateness of the above-referenced procedures. Some of this Protocol may not pertain to the patients you provide care to, as it may relate to products that are not available in your geographic area.

References

We are not responsible for the continuing viability of web site addresses that may be listed in any references below.

- 1. Food and Drug Administration (FDA). Special 510(k): Device Modification 2010. Available online at: http://www.accessdata.fda.gov/cdrh_docs/reviews/K101911.pdf. Last accessed October 2012.
- 2. Ling X, Xiong J, Liang W et al. Can immune cell function assay identify patients at risk of infection or rejection? A meta-analysis. Transplantation 2012; 93(7):737-43.
- 3. Rodrigo E, Lopez-Hoyos M, Corral M et al. ImmuKnow((R)) as a diagnostic tool for predicting infection and acute rejection in adult liver transplant recipients: Systematic review and meta-analysis. Liver Transpl 2012; 18(10):1244-52.
- 4. Kowalski RJ, Post DR, Mannon RB et al. Assessing relative risks of infection and rejection: a meta-analysis using an immune function assay. Transplantation 2006; 82(5):663-8.
- 5. Hooper E, Hawkins DM, Kowalski RJ et al. Establishing pediatric immune response zones using the Cylex ImmuKnow assay. Clin Transplant 2005; 19(6):834-9.
- 6. Kowalski R, Post D, Schneider MC et al. Immune cell function testing: an adjunct to therapeutic drug monitoring in transplant patient management. Clin Transplant 2003; 17(2):77-88.

- 7. Zeevi A, Britz JA, Bentlejewski CA et al. Monitoring immune function during tacrolimus tapering in small bowel transplant recipients. Transpl Immunol 2005; 15(1):17-24.
- 8. Rossano JW, Denfield SW, Kim JJ et al. Assessment of the Cylex ImmuKnow cell function assay in pediatric heart transplant patients. J Heart Lung Transplant 2009; 28(1):26-31.
- 9. Gupta S, Mitchell JD, Markham DW et al. Utility of the Cylex assay in cardiac transplant recipients. J Heart Lung Transplant 2008; 27(8):817-22.
- 10. Shearer G, Clerici M. In vitro analysis of cell-mediated immunity: clinical relevance. Clin Chem 1994; 40(11):2162-65.
- 11. Israeli M, Ben-Gal T, Yaari V et al. Individualized immune monitoring of cardiac transplant recipients by noninvasive longitudinal cellular immunity tests. Transplantation 2010; 89(8):968-76.
- 12. Cabrera R, Ararat M, Soldevila-Pico C et al. Using an immune functional assay to differentiate acute cellular rejection from recurrent hepatitis C in liver transplant patients. Liver Transpl 2009; 15(2):216-22.
- 13. Torio A, Fernandez EJ, Montes-Ares O et al. Lack of association of immune cell function test with rejection in kidney transplantation. Transplant Proc 2011; 43(6):2168-70.
- 14. Heikal NM, Bader FM, Martins TB et al. Immune function surveillance: association with rejection, infection and cardiac allograft vasculopathy. Transplant Proc 2013; 45(1):376-82.
- 15. Serrano M, Meneu JC, Medina E et al. Clinical value of a single determination of intracellular ATP levels in stimulated CD4+ T lymphocytes in pediatric patients with stable liver transplantation. Transplant Proc 2012; 44(9):2622-4.
- 16. Te HS, Dasgupta KA, Cao D et al. Use of immune function test in monitoring immunosuppression in liver transplant recipients. Clin Transplant 2012; 26(6):826-32.
- 17. Zhou H, Lin J, Chen S et al. Use of the ImmuKnow assay to evaluate the effect of alemtuzumab-depleting induction therapy on cell-mediated immune function after renal transplantation. Clin Exp Nephrol 2013; 17(2):304-9.
- 18. Bhorade SM, Janata K, Vigneswaran WT et al. Cylex ImmuKnow assay levels are lower in lung transplant recipients with infection. J Heart Lung Transplant 2008; 27(9):990-4.
- 19. Shino MY, Weigt SS, Saggar R et al. Usefulness of immune monitoring in lung transplantation using adenosine triphosphate production in activated lymphocytes. J Heart Lung Transplant 2012; 31(9):996-1002.
- 20. Reinsmoen NL, Cornett KM, Kloehn R et al. Pretransplant donor-specific and non-specific immune parameters associated with early acute rejection. Transplantation 2008; 85(3):462-70.
- 21. Serban G, Whittaker V, Fan J et al. Significance of immune cell function monitoring in renal transplantation after Thymoglobulin induction therapy. Hum Immunol 2009; 70(11):882-90.
- 22. Husain S, Raza K, Pilewski JM et al. Experience with immune monitoring in lung transplant recipients: correlation of low immune function with infection. Transplantation 2009; 87(12):1852-7.
- 23. Zhou H, Wu Z, Ma L et al. Assessing immunologic function through CD4 T-lymphocyte ahenosine triphosphate levels by ImmuKnow assay in Chinese patients following renal transplantation. Transplant Proc 2011; 43(7):2574-8.
- 24. Kobashigawa JA, Kiyosaki KK, Patel JK et al. Benefit of immune monitoring in heart transplant patients using ATP production in activated lymphocytes. J Heart Lung Transplant 2010; 29(5):504-8.

- 25. Huskey J, Gralla J, Wiseman AC. Single time point immune function assay (ImmuKnow) testing does not aid in the prediction of future opportunistic infections or acute rejection. Clin J Am Soc Nephrol 2011; 6(2):423-9.
- 26. Hashimoto K, Miller C, Hirose K et al. Measurement of CD4+ T-cell function in predicting allograft rejection and recurrent hepatitis C after liver transplantation. Clin Transplant 2010; 24(5):701-8.
- 27. Cheng JW, Shi YH, Fan J et al. An immune function assay predicts post-transplant recurrence in patients with hepatocellular carcinoma. J Cancer Res Clin Oncol 2011; 137(10):1445-53.
- 28. Dong JY, Yin H, Li RD et al. The relationship between adenosine triphosphate within CD4(+) T lymphocytes and acute rejection after liver transplantation. Clin Transplant 2011; 25(3):E292-6.
- 29. Manga K, Serban G, Schwartz J et al. Increased adenosine triphosphate production by peripheral blood CD4+ cells in patients with hematologic malignancies treated with stem cell mobilization agents. Hum Immunol 2010; 71(7):652-8.
- 30. Gesundheit B, Budowski E, Israeli M et al. Assessment of CD4 T-lymphocyte reactivity by the Cylex ImmuKnow assay in patients following allogeneic hematopoietic SCT. Bone Marrow Transplant 2010; 45(3):527-33.
- 31. Kotton CN, Kumar D, Caliendo AM et al. International consensus guidelines on the management of cytomegalovirus in solid organ transplantation. Transplantation 2010; 89(7):779-95.
- 32. Guidelines for the Care of Heart Transplant Recipients, 2010. The International Society of Heart and Lung Transplantation http://www.ishlt.org/publications/guidelines.asp. Accessed October 2011.
- 33. Educational guidelines. American Society of Transplantation. http://www.a-s-t.org/content/educational-guidelines. Accessed October 2011.
- 34. Humar A, Michaels M. American Society of Transplantation recommendations for screening, monitoring and reporting of infectious complications in immunosuppression trials in recipients of organ transplantation. Am J Transplant 2006; 6(2):262-74.